# Package 'evola'

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<b>Description</b> Runs a genetic algorithm using the 'AlphaSimR' machinery <doi:10.1093 g3journal="" jkaa017=""> and the coalescent simulator 'MaCS' <doi:10.1101 gr.083634.108="">.</doi:10.1101></doi:10.1093>
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evola-package

**EVOL**utionary **A**lgorithm

## **Description**

The evola package is nice wrapper of the AlphaSimR package that enables the use of the evolutionary algorithm to solve complex questions in a simple form.

The evolafit function is the core function of the package which allows the user to specify the problem and constraints to find a close-to-optimal solution using the evolutionary forces.

# Keeping evola updated

The evola package is updated on CRAN every 4-months due to CRAN policies but you can find the latest source at https://github.com/covaruber/evola. This can be easily installed typing the following in the R console:

library(devtools)

install\_github("covaruber/evola")

This is recommended if you reported a bug, was fixed and was immediately pushed to GitHub but not in CRAN until the next update.

# **Tutorials**

For tutorials on how to perform different analysis with evola please look at the vignettes by typing in the terminal:

vignette("evola.intro")

# **Getting started**

The package has been equiped with several datasets to learn how to use the evola package:

- \* DT\_technow dataset to perform optimal cross selection.
- \* DT\_wheat dataset to perform optimal training population selection.
- \* DT\_cpdata dataset to perform optimal individual.

#### Models Enabled

The machinery behind the scenes is AlphaSimR.

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#### **Bug report and contact**

If you have any questions or suggestions please post it in https://stackoverflow.com or https://stats.stackexchange.com

I'll be glad to help or answer any question. I have spent a valuable amount of time developing this package. Please cite this package in your publication. Type 'citation("evola")' to know how to cite it

# Author(s)

Giovanny Covarrubias-Pazaran

#### References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. G3 GenelGenomeslGenetics 11(2):jkaa017. https://doi.org/10.1093/g3journal/jkaa017.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. Genome Research, 19, 136-142. http://genome.cshlp.org/content/19/1/136.

A.mat

Additive relationship matrix

# **Description**

Calculates the realized additive relationship matrix.

#### Usage

```
A.mat(X,min.MAF=NULL)
```

#### **Arguments**

X Matrix  $(n \times m)$  of unphased genotypes for n lines and m biallelic markers, coded as  $\{-1,0,1\}$ . Fractional (imputed) and missing values (NA) are allowed.

min.MAF Minimum minor allele frequency. The A matrix is not sensitive to rare alleles,

so by default only monomorphic markers are removed.

#### **Details**

For vanraden method: the marker matrix is centered by subtracting column means M = X - ms where ms is the coumn means. Then A = MM'/c, where  $c = \sum_k d_k/k$ , the mean value of the diagonal values of the MM' portion.

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#### Value

If return.imputed = FALSE, the  $n \times n$  additive relationship matrix is returned. If return.imputed = TRUE, the function returns a list containing

\$A the A matrix

#### References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

# See Also

```
evolafit - the core function of the package
```

## **Examples**

```
## random population of 200 lines with 1000 markers
X <- matrix(rep(0,200*1000),200,1000)
for (i in 1:200) {
    X[i,] <- ifelse(runif(1000)<0.5,-1,1)
}

A <- A.mat(X)

## take a look at the Genomic relationship matrix
    colfunc <- colorRampPalette(c("steelblue4", "springgreen", "yellow"))
    hv <- heatmap(A[1:15,1:15], col = colfunc(100),Colv = "Rowv")
    str(hv)</pre>
```

bestSol

Extract the index of the best solution

#### **Description**

Extracts the index of the best solution for all traits under the constraints specified.

#### Usage

```
bestSol(object, selectTop=TRUE)
```

#### **Arguments**

object A resulting object from the function evolafit.

selectTop Selects highest values for the fitness value if TRUE. Selects lowest values if

FALSE.

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#### **Details**

A simple apply function looking at the fitness value of all the solution in the last generation to find the maximum value.

#### Value

\$res the vector of best solutions in M for each trait in the problem

#### References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

#### See Also

evolafit – the core function of the package

```
set.seed(1)
# Data
Gems <- data.frame(</pre>
  Color = c("Red", "Blue", "Purple", "Orange",
            "Green", "Pink", "White", "Black",
            "Yellow"),
  Weight = round(runif(9, 0.5, 5), 2),
  Value = round(abs(rnorm(9,0,5))+0.5,2),
  Times=c(rep(1,8),0)
head(Gems)
# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.
res0<-evolafit(cbind(Weight, Value)~Color, dt= Gems,</pre>
               # constraints: if greater than this ignore
               constraintsUB = c(10,Inf),
               # constraints: if smaller than this ignore
               constraintsLB= c(-Inf,-Inf),
               # weight the traits for the selection
               traitWeight = c(0,1),
               # population parameters
               nCrosses = 100, nProgeny = 20, recombGens = 1,
               # coancestry parameters
               A=NULL, lambda=c(0,0), nQTLperInd = 1,
               # selection parameters
               propSelBetween = .9, propSelWithin =0.9,
               nGenerations = 50
)
```

DT\_cpdata

bestSol(res0)

DT\_cpdata

Genotypic and Phenotypic data for a CP population

# **Description**

A CP population or F1 cross is the designation for a cross between 2 highly heterozygote individuals; i.e. humans, fruit crops, bredding populations in recurrent selection.

This dataset contains phenotpic data for 363 siblings for an F1 cross. These are averages over 2 environments evaluated for 4 traits; color, yield, fruit average weight, and firmness. The columns in the CPgeno file are the markers whereas the rows are the individuals. The CPpheno data frame contains the measurements for the 363 siblings, and as mentioned before are averages over 2 environments.

#### Usage

```
data("DT_cpdata")
```

#### **Format**

The format is: chr "DT\_cpdata"

#### **Source**

This data was simulated for fruit breeding applications.

# References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. G3 GenelGenomeslGenetics 11(2):jkaa017. https://doi.org/10.1093/g3journal/jkaa017.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. Genome Research, 19, 136-142. http://genome.cshlp.org/content/19/1/136.

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```
# constraints: if sum is smaller than this ignore
              constraintsLB= c(-Inf,-Inf),
              # weight the traits for the selection
              traitWeight = c(1,0),
              # population parameters
              nCrosses = 100, nProgeny = 10,
              recombGens=1, nChr=1, mutRate=0,
              # coancestry parameters
              A=A, lambda= (30*pi)/180, nQTLperInd = 2,
              # selection parameters
              propSelBetween = 0.5, propSelWithin =0.5,
              nGenerations = 40, keepBest=FALSE)
best = bestSol(res)["pop","Yield"];best
xa = (res$M %*% DT$Yield)[best,]; xa
xAx = res$M[best,] %*% A %*% res$M[best,]; xAx
sum(res$M[best,]) # total # of inds selected
pmonitor(res)
plot(DT$Yield, col=as.factor(res$M[best,]),
     pch=(res$M[best,]*19)+1)
pareto(res)
```

DT\_technow

Genotypic and Phenotypic data from single cross hybrids (Technow et al.,2014)

# Description

This dataset contains phenotpic data for 2 traits measured in 1254 single cross hybrids coming from the cross of Flint x Dent heterotic groups. In addition contains the genotipic data (35,478 markers) for each of the 123 Dent lines and 86 Flint lines. The purpose of this data is to demosntrate the prediction of unrealized crosses (9324 unrealized crosses, 1254 evaluated, total 10578 single crosses). We have added the additive relationship matrix (A) but can be easily obtained using the A.mat function on the marker data. Please if using this data for your own research cite Technow et al. (2014) publication (see References).

# Usage

```
data("DT_technow")
```

# **Format**

The format is: chr "DT\_technow"

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#### Source

This data was extracted from Technow et al. (2014).

#### References

If using this data for your own research please cite:

Technow et al. 2014. Genome properties and prospects of genomic predictions of hybrid performance in a Breeding program of maize. Genetics 197:1343-1355.

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. G3 GenelGenomeslGenetics 11(2):jkaa017. https://doi.org/10.1093/g3journal/jkaa017.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. Genome Research, 19, 136-142. http://genome.cshlp.org/content/19/1/136.

```
data(DT_technow)
DT <- DT_technow
DT$occ <- 1; DT$occ[1]=0</pre>
M <- M_technow
  A \leftarrow A.mat(M)
  # run the genetic algorithm
  # we assig a weight to x'Ax of (20*pi)/180=0.34
  res<-evolafit(formula = c(GY, occ)~hy,
                dt = DT,
                # constraints: if sum is greater than this ignore
                constraintsUB = c(Inf, 100),
                # constraints: if sum is smaller than this ignore
                constraintsLB= c(-Inf,-Inf),
                # weight the traits for the selection
                traitWeight = c(1,0),
                # population parameters
                nCrosses = 100, nProgeny = 10,
                recombGens=1, nChr=1, mutRate=0,
                # coancestry parameters
                A=A, lambda= (20*pi)/180, nQTLperInd = 70,
                # selection parameters
                propSelBetween = 0.5, propSelWithin =0.5,
                nGenerations = 20, keepBest=FALSE)
  best = bestSol(res)["pop","GY"]
  xa = (res$M %*% DT$GY)[best,]; xa
  xAx = res$M[best,] %*% A %*% res$M[best,]; xAx
  sum(res$M[best,]) # total # of inds selected
  pmonitor(res)
  plot(DT$GY, col=as.factor(res$M[best,]),
```

DT\_wheat

```
pch=(res$M[best,]*19)+1)
pareto(res)
```

DT\_wheat

wheat lines dataset

#### **Description**

Information from a collection of 599 historical CIMMYT wheat lines. The wheat data set is from CIMMYT's Global Wheat Program. Historically, this program has conducted numerous international trials across a wide variety of wheat-producing environments. The environments represented in these trials were grouped into four basic target sets of environments comprising four main agroclimatic regions previously defined and widely used by CIMMYT's Global Wheat Breeding Program. The phenotypic trait considered here was the average grain yield (GY) of the 599 wheat lines evaluated in each of these four mega-environments.

A pedigree tracing back many generations was available, and the Browse application of the International Crop Information System (ICIS), as described in (McLaren *et al.* 2000, 2005) was used for deriving the relationship matrix A among the 599 lines; it accounts for selection and inbreeding.

Wheat lines were recently genotyped using 1447 Diversity Array Technology (DArT) generated by Triticarte Pty. Ltd. (Canberra, Australia; http://www.triticarte.com.au). The DArT markers may take on two values, denoted by their presence or absence. Markers with a minor allele frequency lower than 0.05 were removed, and missing genotypes were imputed with samples from the marginal distribution of marker genotypes, that is,  $x_{ij} = Bernoulli(\hat{p}_j)$ , where  $\hat{p}_j$  is the estimated allele frequency computed from the non-missing genotypes. The number of DArT MMs after edition was 1279.

#### Usage

```
data(DT_wheat)
```

#### **Format**

Matrix Y contains the average grain yield, column 1: Grain yield for environment 1 and so on.

#### **Source**

International Maize and Wheat Improvement Center (CIMMYT), Mexico.

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#### References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. G3 GenelGenomeslGenetics 11(2):jkaa017. https://doi.org/10.1093/g3journal/jkaa017.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. Genome Research, 19, 136-142. http://genome.cshlp.org/content/19/1/136.

McLaren, C. G., L. Ramos, C. Lopez, and W. Eusebio. 2000. "Applications of the geneaology manegment system." In *International Crop Information System. Technical Development Manual, version VI*, edited by McLaren, C. G., J.W. White and P.N. Fox. pp. 5.8-5.13. CIMMyT, Mexico: CIMMyT and IRRI.

McLaren, C. G., R. Bruskiewich, A.M. Portugal, and A.B. Cosico. 2005. The International Rice Information System. A platform for meta-analysis of rice crop data. *Plant Physiology* **139**: 637-642.

```
# example to optimize a training pop for a validation pop
data(DT_wheat)
DT <- as.data.frame(DT_wheat)</pre>
DT$id <- rownames(DT) # IDs
DT$occ <- 1; DT$occ[1]=0 # to track occurrences
DT$dummy <- 1; DT$dummy[1]=0 # dummy trait
# if genomic
GT <- GT_wheat + 1; rownames(GT) <- rownames(DT)
G <- GT%*%t(GT)
G <- G/mean(diag(G))</pre>
# if pedigree
A <- A_wheat
A[1:4,1:4]
##Perform eigenvalue decomposition for clustering
##And select cluster 5 as target set to predict
pcNum=25
svdWheat <- svd(A, nu = pcNum, nv = pcNum)</pre>
PCWheat <- A %*% svdWheat$v
rownames(PCWheat) <- rownames(A)</pre>
DistWheat <- dist(PCWheat)</pre>
TreeWheat <- cutree(hclust(DistWheat), k = 5 )</pre>
plot(PCWheat[,1], PCWheat[,2], col = TreeWheat,
     pch = as.character(TreeWheat), xlab = "pc1", ylab = "pc2")
vp <- rownames(PCWheat)[TreeWheat == 3]; length(vp)</pre>
tp <- setdiff(rownames(PCWheat),vp)</pre>
As <- A[tp,tp]
DT2 <- DT[rownames(As),]</pre>
DT2$cov <- apply(A[tp,vp],1,mean)
head(DT2)
```

```
# we assign a weight to x'Ax of (60*pi)/180=1
res<-evolafit(formula=cbind(cov, occ)~id, dt= DT2,</pre>
              # constraints: if sum is greater than this ignore
              constraintsUB = c(Inf, 100),
              # constraints: if sum is smaller than this ignore
              constraintsLB= c(-Inf, -Inf),
              # weight the traits for the selection
              traitWeight = c(1,0),
              # population parameters
              nCrosses = 100, nProgeny = 10,
              recombGens=1, nChr=1, mutRate=0,
              # coancestry parameters
              A=As, lambda= (60*pi)/180, nQTLperInd = 80,
              # selection parameters
              propSelBetween = 0.5, propSelWithin =0.5,
              nGenerations = 30, verbose = TRUE, keepBest=FALSE)
best <- bestSol(res)["pop","cov"]</pre>
sum(res$M[best,]) # total # of inds selected
pareto(res)
```

evolafit

Fits a genetic algorithm for a set of traits and constraints.

# **Description**

Using the AlphaSimR machinery it recreates the evolutionary forces applied to a problem where possible solutions replace individuals and combinations of variables to optimize in the problem replace the genes or QTLs. Then evolutionary forces (mutation, selection and drift) are applied to find a close-to-optimal solution.

# Usage

# **Arguments**

formula Formula of the form y~x where 'y' refers to the traits or features defining the

average allelic substitution effects of the QTLs, and 'x'refers to the variable

defining the genes or QTLs to be combined in the possible solutions.

A dataset containing the features/traits and classifiers/genes/QTLs.

constraintsUB A numeric vector specifying the upper bound constraints in the traits/features

> (y). The length is equal to the number of traits/features in the formula. If missing is assume an infinite value for all traits. Solutions with higher value than the upper bound are assigned a -infinite value if the argument selectTop=TRUE and to +infinite when selectTop=FALSE, which is equivalent to reject/discard a

solution based on the fitness function.

constraintsLB A numeric vector specifying the lower bound constraints in the traits/features

> (y). The length is equal to the number of traits/features in the formula. If missing is assume a -infinite value for all traits. Solutions with lower value than the lower bound are assigned a +infinite value if the argument selectTop=TRUE and to infinite when selectTop=FALSE, which is equivalent to reject/discard a solution

based on the fitness function.

traitWeight A numeric vector specifying the weights that each trait has in the fitness function

(e.g., a selection index). The length is equal to the number of traits/features. If

missing is assumed equal weight (1) for all traits.

nCrosses A numeric value indicating how many crosses should occur in the population of

solutions at every generation.

A numeric value indicating how many progeny (solutions) each cross should nProgeny

generate in the population of solutions at every generation.

The number of generations that the evolutionary process should run for. nGenerations

recombGens The number of recombination generations that should occur before selection is

applied. This is in case the user wants to allow for more recombination before

selection operates. The default is 1.

nChr The number of chromosomes where features/genes should be allocated to. The

default value is 1 but this number can be increased to mimic more recombination

events at every generation. STILL NOT FUNCTIONAL.

mutRate A value between 0 and 1 to indicate the proportion of random QTLs that should

> mutate in each individual. For example, a value of 0.1 means that a random 10% of the QTLs will mutate in each individual randomly taking values of 0 or 1. Is important to notice that this implies that the stopping criteria based in variance will never be reached because we keep introducing variance through random

mutation.

nOTLperInd The number of OTLs/genes (classifier x) that should be fixed for the positive

allele at the begginning of the simulation. If not specified it will be equal to the

20% of the QTLs (number of rows in the dt over 5). See details section.

A relationship matrix between the levels of the classifier variable (x or QTLs;

not between the solutions). It is a kind of a linkage disequilibrium matrix. This

function can be used or ignored in the fitness function.

dt

Α

lambda A numeric value indicating the weight assigned to the relationship between lev-

els of the classifiers in comparison with the trait value. If not specified is assumed to be 0. This can be used or ignored in your customized fitness function.

propSelBetween A numeric value between 0 and 1 indicating the proportion of families/crosses

that should be selected. The default is 1, meaning all crosses are selected.

propSelWithin A numeric value between 0 and 1 indicating the proportion of individuals within

families/crosses that should be selected. The default value is 0.5, meaning that

50% of the top individuals are selected.

fitnessf An alternative fitness function for a linear combination of the traits. If NULL

the default function will be: function $(Y,b,d,Q)\{(Y\%*\%b) - d\}$ 

where Y%\*%b is equivalent to xa in contribution theory, and d is equal to x'Ax, being x the contribution vector to the solution, a are the QTL effects, and A is the covariance between QTLs, Q is the QTL matrix for the solution. If you provide your own fitness function please keep in mind that the variables Y, b, d

and Q are already reserved and should always be added to your function (even

if not used) in addition to your new variables.

verbose A logical value indicating if we should print logs.

dateWarning A logical value indicating if you should be warned when there is a new version

on CRAN.

selectTop Selects highest values for the fitness value if TRUE. Selects lowest values if

FALSE.

tolVarG A stopping criteria (tolerance for genetic variance) when the variance across

traits is smaller than this value, which is equivalent to assume that all solutions having the same QTL profile (depleted variance). The default value is 1e-6 and is computed as the sum of the diagonal values of the genetic variance covariance

matrix between traits.

keepBest A TRUE/FALSE value to indicate if we should store the QTL matrix and pedi-

gree of the solutions selected across generations. This can be useful if we want to recreate the path to the best solution (e.g., best crossing path to a product).

... Further arguments to be passed to the fitness function.

#### **Details**

Using the AlphaSimR machinery (runMacs) it recreates the evolutionary forces applied to a problem where possible solutions replace individuals and combinations of variables in the problem replace the genes. Then evolutionary forces are applied to find a close-to-optimal solution. The number of solutions are controlled with the nCrosses and nProgeny parameters, whereas the number of initial QTLs activated in a solution is controlled by the nQTLperInd parameter. The number of activated QTLs of course will increase if has a positive effect in the fitness of the solutions. The drift force can be controlled by the recombGens parameter. The mutation rate can be controlled with the mutRate parameter. The recombination rate can be controlled with the nChr argument.

#### Value

**\$M** the matrix of haplotypes/solutions at the end of the run.

\$Mb the matrix of top (parents) haplotypes/solutions at the end of the run.

**\$score** a matrix with scores for different metrics across n generations of evolution.

**\$pheno** the matrix of phenotypes of individuals/solutions present in the last generation.

**\$phenoBest** the matrix of phenotypes of top (parents) individuals/solutions present in the last generation.

indivPerformance the matrix of x'a, x'Ax, deltaC, nQTLs per solution per generation.

**pop** AlphaSimR object used for the evolutionary algorithm at the last iteration.

**best** AlphaSimR object corresponding to the best parental haplotypes/solutions selected for new crosses across all cycles.

**pedBest** if the argument keepBest=TRUE this contains the pedigree of the selected solutions across iterations.

**traits** a character vector corresponding to the name of the variables used in the fitness function.

#### References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

Gaynor, R. Chris, Gregor Gorjanc, and John M. Hickey. 2021. AlphaSimR: an R package for breeding program simulations. G3 GenelGenomeslGenetics 11(2):jkaa017. https://doi.org/10.1093/g3journal/jkaa017.

Chen GK, Marjoram P, Wall JD (2009). Fast and Flexible Simulation of DNA Sequence Data. Genome Research, 19, 136-142. http://genome.cshlp.org/content/19/1/136.

#### See Also

evolafit – the information of the package

```
set.seed(1)
# Data
Gems <- data.frame(</pre>
 Color = c("Red", "Blue", "Purple", "Orange",
            "Green", "Pink", "White", "Black",
            "Yellow"),
 Weight = round(runif(9, 0.5, 5), 2),
 Value = round(abs(rnorm(9,0,5))+0.5,2),
 Times=c(rep(1,8),0)
)
head(Gems)
     Color Weight Value
      Red 4.88 9.95
     Blue
            1.43 2.73
# 3 Purple
            1.52 2.60
# 4 Orange
            3.11 0.61
# 5 Green
            2.49 0.77
     Pink
            3.53 1.99
            0.62 9.64
    White
```

*Jc* 15

```
# 8 Black 2.59 1.14
# 9 Yellow
            1.77 10.21
# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.
res0<-evolafit(formula=cbind(Weight, Value)~Color, dt= Gems,</pre>
               # constraints: if greater than this ignore
               constraintsUB = c(10,Inf),
               # constraints: if smaller than this ignore
               constraintsLB= c(-Inf,-Inf),
               # weight the traits for the selection
               traitWeight = c(0,1),
               # population parameters
               nCrosses = 100, nProgeny = 20,
               # genome parameters
               recombGens = 1, nChr=1, mutRate=0, nQTLperInd = 1,
               # coancestry parameters
               A=NULL, lambda=0,
               # selection parameters
               propSelBetween = .9, propSelWithin =0.9,
               nGenerations = 50
)
best = bestSol(res0)["pop","Value"]
xa = res0$M[best,] %*% as.matrix(Gems[,c("Weight","Value")]); xa
res0$M[best,]
res0$score[nrow(res0$score),]
# $`Genes`
# Red Blue Purple Orange Green Pink White Black Yellow
                                     0
# $Result
# Weight Value
# 8.74 32.10
pmonitor(res0)
pareto(res0)
```

Jc

Matrix of ones

#### **Description**

Makes a matrix of ones with a single row and nc columns.

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# Usage

Jc(nc)

# **Arguments**

nc

Number of columns to create.

# **Details**

A simple apply function to make a matrix of one row and nc columns.

# Value

\$res a matrix

# References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

# See Also

```
evolafit – the core function of the package
```

# **Examples**

Jc(5)

Jr

Matrix of ones

# Description

Makes a matrix of ones with a single column and nr rows.

# Usage

Jr(nr)

# **Arguments**

nr

Number of rows to create.

#### **Details**

A simple apply function to make a matrix of one column and nr rows.

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#### Value

\$res a matrix

#### References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

#### See Also

```
evolafit – the core function of the package
```

#### **Examples**

Jr(5)

overlay

Overlay Matrix

# **Description**

'overlay' adds r times the design matrix for model term t to the existing design matrix. Specifically, if the model up to this point has p effects and t has a effects, the a columns of the design matrix for t are multiplied by the scalar r (default value 1.0). This can be used to force a correlation of 1 between two terms as in a diallel analysis.

#### Usage

```
overlay(..., rlist=NULL, prefix=NULL, sparse=FALSE)
```

# **Arguments**

as many vectors as desired to overlay. rlist a list of scalar values indicating the times that each incidence matrix overlayed should be multiplied by. By default r=1. a character name to be added before the column names of the final overlay maprefix trix. This may be useful if you have entries with names starting with numbers which programs such as asreml doesn't like, or for posterior extraction of parameters, that way 'grep'ing is easier. sparse

a TRUE/FALSE statement specifying if the matrices should be built as sparse or

regular matrices.

#### Value

\$\$3 an incidence matrix with as many columns levels in the vectors provided to build the incidence matrix.

18 pareto

#### Author(s)

Giovanny Covarrubias-Pazaran

#### References

Fikret Isik. 2009. Analysis of Diallel Mating Designs. North Carolina State University, Raleigh, USA.

Covarrubias-Pazaran G (2016) Genome assisted prediction of quantitative traits using the R package soevolafit. PLoS ONE 11(6): doi:10.1371/journal.pone.0156744

#### See Also

The core functions of the package evolafit.

# **Examples**

pareto

plot the change of values across iterations

# Description

```
plot for monitoring.
```

#### Usage

```
pareto(object, scaled=TRUE,pch=20, xlim, ...)
```

#### **Arguments**

object	model object of class "evolafit"
scaled	a logical value to specify the scale of the y-axis (gain in merit).
pch	symbol for plotting points as desribed in par
xlim	upper and lower bound in the x-axis
	Further arguments to be passed to the plot function.

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# Value

vector of plot

# Author(s)

Giovanny Covarrubias

# See Also

```
plot, evolafit
```

 ${\tt pmonitor}$ 

plot the change of values across iterations

# Description

```
plot for monitoring.
```

# Usage

```
pmonitor(object, ...)
```

# Arguments

object model object of class "evolafit"

... Further arguments to be passed to the plot function.

# Value

vector of plot

# Author(s)

Giovanny Covarrubias

# See Also

```
plot, evolafit
```

20 stan

stan

Standardize a vector of values in range 0 to 1

# Description

Simple function to map a vector of values to the range of 0 and 1 values to have a better behavior of the algorithm.

# Usage

```
stan(x)
```

# **Arguments**

Χ

A vector of numeric values.

# **Details**

Simple function to map a vector of values to the range of 0 and 1 values to have a better behavior of the algorithm.

# Value

**\$res** new values in range 0 to 1

# References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

# See Also

```
evolafit – the core function of the package
```

```
x <- rnorm(20, 10, 3);x
stan(x)</pre>
```

varM 21

varM

Extract the variance existing in the genome solutions

# Description

Extracts the variance found across the M element of the resulting object of the evolafit() function which contains the different solution and somehow represents the genome of the population.

# Usage

```
varM(object)
```

# Arguments

object

A resulting object from the function evolafit.

#### **Details**

A simple apply function looking at the variance in each column of the M element of the resulting object of the evolafit function.

# Value

**\$res** a value of variance

#### References

Giovanny Covarrubias-Pazaran (2024). evola: a simple evolutionary algorithm for complex problems. To be submitted to Bioinformatics.

# See Also

```
evolafit – the core function of the package
```

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```
# Task: Gem selection.
# Aim: Get highest combined value.
# Restriction: Max weight of the gem combined = 10.
res0<-evolafit(cbind(Weight, Value)~Color, dt= Gems,</pre>
               # constraints: if greater than this ignore
               constraintsUB = c(10,Inf),
               # constraints: if smaller than this ignore
               constraintsLB= c(-Inf,-Inf),
               # weight the traits for the selection
               traitWeight = c(0,1),
               # population parameters
               nCrosses = 100, nProgeny = 20, recombGens = 1,
               # coancestry parameters
               A=NULL, lambda=c(0,0), nQTLperInd = 1,
               # selection parameters
               propSelBetween = .9, propSelWithin =0.9,
               nGenerations = 50
)
varM(res0)
```

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