

# Package ‘Uniquorn’

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**Title** Identification of cancer cell lines based on their weighted mutational/ variational fingerprint

**Version** 1.0.8

**Description** This packages enables users to identify cancer cell lines. Cancer cell line misidentification and cross-contamination reprints a significant challenge for cancer researchers. The identification is vital and in the frame of this package based on the locations/ loci of somatic and germline mutations/ variations. The input format is vcf/ vcf.gz and the files have to contain a single cancer cell line sample (i.e. a single member/genotype/gt column in the vcf file). The implemented method is optimized for the Next-generation whole exome and whole genome DNA-sequencing technology.

**Imports** DBI, stringr, RSQLite, R.utils, WriteXLS

**Depends** R (>= 3.3)

**License** Artistic-2.0

**LazyData** TRUE

**Type** Package

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**Suggests** testthat, knitr, rmarkdown, BiocGenerics, RUnit

**biocViews** Software, StatisticalMethod, WholeGenome

**VignetteBuilder** knitr

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add\_custom\_vcf\_to\_database

*Adds a custom vcf file to the three existing cancer cell line panels*

---

### Description

Adds a custom vcf file to the three existing cancer cell line panels

### Usage

```
add_custom_vcf_to_database(
    vcf_file_path,
    ref_gen = "GRCH37",
    name_cl = "",
    safe_mode = FALSE,
    test_mode = FALSE)
```

### Arguments

vcf_file_path	Input vcf file. Only one sample column allowed.
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
name_cl	Name of the to-be-added cancer cell line sample. '_CUSTOM' will automatically be added as suffix.
safe_mode	Only add mutations to the database where there already are mutations found in the canonical cancer cell lines. This is a safety mechanism against overfitting if there are too few custom training samples.

test\_mode      Is this a test? Just for internal use

**Value**

Message if the adding has succeeded

**Examples**

```
HT29_vcf_file = system.file("extdata/HT29.vcf.gz", package="Uniquorn");
add_custom_vcf_to_database(
vcf_file_path = HT29_vcf_file,
name_cl = "",
ref_gen = "GRCH37",
safe_mode = FALSE,
test_mode = TRUE )
```

---

add\_missing\_cls      *add\_missing\_cls*

---

**Description**

add\_missing\_cls

**Usage**

```
add_missing_cls(res_table, dif_cls)
```

**Arguments**

res\_table      Table that contains the identification results  
dif\_cls      Missing CLs

**Value**

Results table with added missing cls

---

calculate\_p\_and\_q\_values  
*calculate\_p\_and\_q\_values*

---

### Description

calculate\_p\_and\_q\_values

### Usage

```
calculate_p_and_q_values(candidate_hits_abs_all, cl_absolute_mutation_hits,  
    sim_list, sim_list_stats, minimum_matching_mutations, list_of_cls, p_value,  
    q_value, vcf_fingerprint, panels)
```

### Arguments

candidate_hits_abs_all	Maximally possible found variants
cl_absolute_mutation_hits	Matching variants
sim_list	Contains reference mutation data
sim_list_stats	Contains global reference mutation stats
minimum_matching_mutations	Minimal amount of required matching mutations
list_of_cls	List of CLs
p_value	Required maximal p-value
q_value	Required maximal q-value
vcf_fingerprint	The start and end positions of variants in the query
panels	The reference libraries

### Value

Results table

---

```
calculate_similarity_results  
    calculate_similarity_results
```

---

## Description

calculate\_similarity\_results

## Usage

```
calculate_similarity_results(sim_list, sim_list_stats, found_mut_mapping,  
    minimum_matching_mutations, p_value, q_value, confidence_score,  
    vcf_fingerprint, panels, list_of_cls)
```

## Arguments

sim_list	Contains reference mutation data
sim_list_stats	Contains global reference mutation stats
found_mut_mapping	Mapping to mutations from query to reference mutation set
minimum_matching_mutations	Minimal amount of required matching mutations
p_value	Required maximal p-value
q_value	Required maximal q-value
confidence_score	Threshold above which a positive prediction occurs default 3.0
vcf_fingerprint	The start and end positions of variants in the query
panels	The reference libraries
list_of_cls	List of cancer cell lines

## Value

Results table

---

create\_bed\_file      *create\_bed\_file*

---

### Description

Creates BED files from the found and not found annotated mutations

### Usage

```
create_bed_file(  
  sim_list,  
  vcf_fingerprint,  
  res_table,  
  output_file,  
  ref_gen,  
  manual_identifier  
)
```

### Arguments

sim_list	R table which contains the mutations from the training database for the cancer cell lines
vcf_fingerprint	contains the mutations that are present in the query cancer cell line's vcf file
res_table	Table containing the identification results
output_file	Path to output file
ref_gen	Reference genome version
manual_identifier	Manually enter a vector of CL name(s) whose bed files should be created, independently from them passing the detection threshold

### Value

Returns a message which indicates if the BED file creation has succeeded

---

filter\_for\_weights     *filter\_for\_weights*

---

### Description

Filter the reference set

### Usage

```
filter_for_weights(  
mutational_weight_inclusion_threshold,  
ref_gen,  
verbose,  
sim_list,  
sim_list_stats)
```

### Arguments

mutational_weight_inclusion_threshold	Lower bound for mutational weight to be included
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
verbose	Print additional information
sim_list	Contains the mutations
sim_list_stats	Contains the overall mutation statistics

### Details

filter\_for\_weights parses vcf file and output basic information

### Value

Filtered reference sets

---

identify\_vcf\_file     *identify\_VCF\_file*

---

### Description

Identifies a cancer cell lines contained in a vcf file based on the pattern (start & length) of all contained mutations/ variations.

**Usage**

```

identify_vcf_file(
vcf_file,
output_file = "",
ref_gen = "GRCH37",
minimum_matching_mutations = 0,
mutational_weight_inclusion_threshold = 1.0,
only_first_candidate = FALSE,
write_xls = FALSE,
output_bed_file = FALSE,
manual_identifier_bed_file = "",
verbose = FALSE,
p_value = .05,
q_value = .05,
confidence_score = 25.0)

```

**Arguments**

vcf_file	Input vcf file. Only one sample column allowed.
output_file	Path of the output file. If blank, autogenerated as name of input file plus '_uniquorn_ident.tab' suffix.
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
minimum_matching_mutations	The minimum amount of mutations that has to match between query and training sample for a positive prediction
mutational_weight_inclusion_threshold	Include only mutations with a weight of at least x. Range: 0.0 to 1.0. 1= unique to CL. ~0 = found in many CL samples.
only_first_candidate	Only the CL identifier with highest score is predicted to be present in the sample
write_xls	Create identification results additionally as xls file for easier reading
output_bed_file	If BED files for IGV visualization should be created for the Cancer Cell lines that pass the threshold
manual_identifier_bed_file	Manually enter a vector of CL name(s) whose bed files should be created, independently from them passing the detection threshold
verbose	Print additional information
p_value	Required p-value for identification
q_value	Required q-value for identification
confidence_score	Threshold above which a positive prediction occurs default 25.0

**Details**

identify\_vcf\_file parses the vcf file and predicts the identity of the sample

**Value**

R table with a statistic of the identification result

**Examples**

```
HT29_vcf_file = system.file("extdata/HT29.vcf.gz", package="Uniquorn");  
identification = identify_vcf_file( HT29_vcf_file )
```

---

```
initiate_canonical_databases  
      initiate_canonical_databases
```

---

**Description**

Parses data into r list variable

**Usage**

```
initiate_canonical_databases(  
  cosmic_file = "CosmicCLP_MutantExport.tsv",  
  ccle_file = "CCLE_hybrid_capture1650_hg19_NoCommonSNPs_CDS_2012.05.07.maf",  
  ref_gen = "GRCH37")
```

**Arguments**

cosmic_file	The path to the cosmic DNA genotype data file. Ensure that the right reference genome is used
ccle_file	The path to the ccle DNA genotype data file. Ensure that the right reference genome is used
ref_gen	Reference genome version

**Value**

Returns message if parsing process has succeeded

**Examples**

```
initiate_canonical_databases(  
  cosmic_file = "CosmicCLP_MutantExport.tsv",  
  ccle_file = "CCLE_hybrid_capture1650_hg19_NoCommonSNPs_CDS_2012.05.07.maf",  
  ref_gen = "GRCH37")
```

---

```
initiate_db_and_load_data
    initiate_db_and_load_data
```

---

**Description**

Intern utility function, loads database and return the `sim_list` and `sim_list_stats` variables.

**Usage**

```
initiate_db_and_load_data(
    ref_gen,
    request_table,
    load_default_db )
```

**Arguments**

<code>ref_gen</code>	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
<code>request_table</code>	Names of the tables to be extracted from the database
<code>load_default_db</code>	Indicate whether the default db should be used as source for the data

**Value**

Returns the `sim_list` and `sim_list_stats` variable

---

```
init_and_load_identification
    init_and_load_identification
```

---

**Description**

Initiate the analysis Output basic information

**Usage**

```
init_and_load_identification(
    verbose,
    ref_gen,
    vcf_file,
    output_file)
```

**Arguments**

<code>verbose</code>	Print additional information
<code>ref_gen</code>	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
<code>vcf_file</code>	Path to <code>vcf_file</code>
<code>output_file</code>	Path to output report file

**Details**

`init_and_load_identification` parses vcf file and output basic information

**Value**

Three file path instances and the fingerprint

---

`parse_ccle_genotype_data`  
*parse\_ccle\_genotype\_data*

---

**Description**

Parses ccle genotype data

**Usage**

```
parse_ccle_genotype_data(ccle_file, sim_list)
```

**Arguments**

<code>ccle_file</code>	Path to CCLE file on hard disk
<code>sim_list</code>	Variable containing mutations and cell line

**Value**

The R Table `sim_list` which contains the CCLE fingerprints

---

parse\_cosmic\_genotype\_data  
*parse\_cosmic\_genotype\_data*

---

**Description**

Parses cosmic genotype data

**Usage**

```
parse_cosmic_genotype_data(cosmic_file, sim_list)
```

**Arguments**

cosmic\_file     Path to cosmic clp file in hard disk  
sim\_list        Variable containing mutations & cell line

**Value**

The R Table sim\_list which contains the CoSMIC CLP fingerprints

---

parse\_vcf\_file     *parse\_vcf\_file*

---

**Description**

Parses the vcf file and filters all information except for the start and length of variations/ mutations.

**Usage**

```
parse_vcf_file( vcf_file_path )
```

**Arguments**

vcf\_file\_path    Path to the vcf file on the operating system

**Value**

Loci-based DNA-mutational fingerprint of the cancer cell line as found in the input VCF file

---

`remove_custom_vcf_from_database`

*Removes a cancer cell line training fingerprint (vcf file) from the database. The names of all training sets can be seen by using the function show\_contained\_cls.*

---

**Description**

Removes a cancer cell line training fingerprint (vcf file) from the database. The names of all training sets can be seen by using the function show\_contained\_cls.

**Usage**

```
remove_custom_vcf_from_database(  
name_cl,  
ref_gen = "GRCH37",  
test_mode = FALSE)
```

**Arguments**

<code>name_cl</code>	name of the cancer cell line training fingerprint
<code>ref_gen</code>	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
<code>test_mode</code>	Is this a test? Just for internal use

**Value**

Message that indicates if the removal was successful

**Examples**

```
remove_custom_vcf_from_database(  
name_cl = "HT29_CELLMINER",  
ref_gen = "GRCH37",  
test_mode = TRUE )
```

---

`re_calculate_cl_weights`

*Re-calculate sim\_list\_weights*

---

**Description**

This function re-calculates the weights of mutation after a change of the training set

**Usage**

```
re_calculate_cl_weights(sim_list, ref_gen)
```

**Arguments**

<code>sim_list</code>	R Table which contains a mapping from mutations/ variations to their containing CLs
<code>ref_gen</code>	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37

**Value**

A list containing both the `sim_list` at pos 1 and `sim_list_stats` at pos 2 data frames.

---

<code>show_contained_cls</code>	<i>show_contained_cls</i>
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---

**Description**

Show all cancer cell line identifier present in the database for a selected reference genome: This function shows the names, amount of mutations/ variations, overall weight of the mutations of all contained training CLs for a chosen reference genome.

**Usage**

```
show_contained_cls(  
  ref_gen)
```

**Arguments**

<code>ref_gen</code>	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
----------------------	---

**Value**

R table which contains the identifier of all cancer cell line samples with the specific reference genome and the weight of all mutations

**Examples**

```
contained_cls = show_contained_cls(  
  ref_gen = "GRCH37")
```

---

```
show_contained_mutations  
    show_contained_mutations
```

---

**Description**

Show all mutations present in the database for a selected reference Genome: This function shows all training-set mutations for a selected reference genome, i.e. the mutations that are being used for identification of query cancer cell lines.

**Usage**

```
show_contained_mutations(  
  ref_gen )
```

**Arguments**

ref\_gen            Reference genome version

**Value**

R Table which contains all mutations associated with a particular cancer cell line for a specified reference genome

**Examples**

```
contained_cls = show_contained_mutations( ref_gen = "GRCH37" )
```

---

```
show_contained_mutations_for_cl  
    show_contained_mutations_for_cl
```

---

**Description**

Show all mutations present in the database for a selected cancer cell line and reference Genome

**Usage**

```
show_contained_mutations_for_cl(  
  name_cl,  
  ref_gen)
```

**Arguments**

name\_cl            Name of the cancer cell line sample stored in the database  
ref\_gen            Reference genome version

**Value**

R table which contains all mutations associated with the defined cancer cell line and reference genome

**Examples**

```
SK_OV_3_CELLMINER_mutations = show_contained_mutations_for_cl(  
  name_cl = "SK_OV_3_CELLMINER_mutations",  
  ref_gen = "GRCH37")
```

---

```
show_which_cls_contain_mutation  
  show_which_cls_contain_mutation
```

---

**Description**

Show all cancer cell lines in the database which contained the specified mutation and reference Genome. Closed interval coordinates. Format mutation: CHR\_START\_STOP, e.g. 1\_123\_123

**Usage**

```
show_which_cls_contain_mutation(  
  mutation_name,  
  ref_gen)
```

**Arguments**

<code>mutation_name</code>	Name of the mutation in the format CHROMOSOME_START_STOP, e.g. '11_244501_244510'
<code>ref_gen</code>	Reference genome version

**Value**

R table which contains all cancer cell line samples which contain the specified mutation with respect to the specified reference genome version

**Examples**

```
Cls_containing_mutations = show_which_cls_contain_mutation(  
  mutation_name = "10_103354427_103354427",  
  ref_gen = "GRCH37")
```

---

split_add	<i>split_add</i>
-----------	------------------

---

**Description**

split\_add

**Usage**

split\_add(vcf\_matrix\_row)

**Arguments**

vcf\_matrix\_row row of the vcf file

**Value**

Transformed entry of vcf file, reduced to start and length

---

write_data_to_db	<i>write_data_to_db</i>
------------------	-------------------------

---

**Description**

Intern utility function, writes to database the sim\_list and sim\_list\_stats variables

**Usage**

```
write_data_to_db(
  content_table,
  table_name,
  ref_gen,
  overwrite,
  test_mode )
```

**Arguments**

content_table	Tables to be written in db
table_name	Name of the table to be written into the DB
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
overwrite	Overwrite the potentially existing table
test_mode	Is this a test? Just for internal use

**Value**

the sim\_list and sim\_list\_stats variable

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